

Article

pubs.acs.org/crystal

¹ Interactions of Aromatic Residues in Amyloids: A Survey of Protein ² Data Bank Crystallographic Data

³ Ivana M. Stanković,[†] Dragana M. Božinovski,[‡] Edward N. Brothers,[§] Milivoj R. Belić,[§] Michael B. Hall,[¶]

s [†]Institute of Chemistry, Technology and Metallurgy, University of Belgrade, Njegoševa 12, 11000 Belgrade, Serbia

6 [‡]Department of Chemistry, University of Belgrade, Studentski trg 12-16, 11000 Belgrade, Serbia

7 [§]Science Program, Texas A&M University at Qatar, Texas A&M Engineering Building, Education City, Doha, Qatar

8 ^{II}Department of Chemistry, Texas A&M University, College Station, Texas 77843-3255, United States

9 **(3)** Supporting Information

10 ABSTRACT: Aromatic-aromatic interactions have long been considered important in the selfassembly of amyloids. In spite of their importance, aromatic amino acids are not detected in every 11 amyloid. In the present study, the occurrence and geometry of these interactions were analyzed 12 for the amyloid structures found in the Protein Data Bank. The data confirm that aromatic amino 13 acids are not crucial for amyloid fibril formation. In fact, aromatic-aliphatic interactions are more 14 frequent than the aromatic-aromatic interactions. Aromatic-aliphatic interactions are present in 15 higher numbers of structures and in certain amyloid sequences; they are more frequent than 16 aromatic-aromatic interactions. An analysis of aromatic/aromatic interactions shows different 17 interaction geometries in intrasheet and intersheet contacts; the intrasheet aromatic-aromatic 18 interactions are mostly parallel and displaced, while intersheet interactions are not parallel. Thus, 19 among the aromatic-aromatic interactions there are important edge-to-face attractions in 20 addition to parallel stacking ones. 21

22 INTRODUCTION

²³ Amyloids are insoluble proteins of a cross- β structure found as 24 deposits in many neurodegenerative diseases, such as 25 Alzheimer's, Parkinson's, Creutzfeldt-Jakob's, Huntington's, ²⁶ or in type II diabetes.^{1–6} Because of their strong fibrillar nature, 27 they can be found in normal tissues as well, like nails, spider 28 webs, or silk.⁷ Amyloids have attracted great attention because 29 of their perceived role in various diseases, unique architecture, 30 and exceptional physical properties.^{6,8} Short polypeptides, with 31 a minimum length of four amino acids, are self-assembled into ₃₂ β -sheets via backbone hydrogen bonds, and then several β -33 sheets interact with each other via polypeptide side chains, to 34 form long linear unbranched protofilaments with an axis nearly 35 perpendicular to a polypeptide strand.⁹ Several protofilaments, 36 the number being specific to the particular amyloid protein, 37 form fibrils.¹⁰ All amyloid proteins, independent of their 38 sequence, form similar structures, namely, the cross- β structure 39 which is made of parallel arrays of β -strands. These structures 40 differ only in the intersheet spacing, which depends on the side 41 chain size, and in the morphology of a fibril.¹⁰

⁴² Although they are not indispensable, the aromatic amino ⁴³ acids phenylalanine (Phe), tyrosine (Tyr), and tryptophan ⁴⁴ (Trp) appear to be important in amyloid formation, kinetics, ⁴⁵ and thermodynamic stability.^{9,11–22} Aromatic amino acids are ⁴⁶ hydrophobic and have a high β -sheet propensity. These ⁴⁷ properties appear crucial in amyloid formation. Furthermore, ⁴⁸ aromatic amino acids possess an ability to engage in π – π interactions and have a directing role in the kinetics of amyloid 49 formation.^{9,10,12,14,15,23} 50

Aromatic–aromatic interactions (Ar/Ar) generally give rise $_{51}$ to three different types of geometries that differ by the angle $_{52}$ between rings and offset values: edge-to-face/T-shaped, face-to- $_{53}$ face, and parallel displaced (offset stacked) interactions (Figure $_{54 \text{ fi}}$ 1). Generally, the face-to-face orientation is rarely observed, as $_{55 \text{ fi}}$ it leads to an unfavorable electrostatic repulsion between the $_{56}$ two planar faces of the aromatic rings. The majority of $_{57}$ interactions in the proteins in general fall into a T-shaped $_{58}$ orientation.^{24–26}



Figure 1. Representation of the three aromatic-aromatic interaction types.

 Received:
 July 25, 2017

 Revised:
 October 19, 2017

 Published:
 October 23, 2017

⁶⁰ Our previous work based on the analysis of crystal structures ⁶¹ from the Cambridge Structural Database (CSD), protein ⁶² structures from the Protein Data Bank (PDB), and quantum ⁶³ chemical calculations showed the importance of parallel ⁶⁴ aromatic–aromatic interactions at large offsets of 4.5-5.5⁶⁵ Å.^{26,27} The quantum chemical calculations indicate that even at ⁶⁶ large offsets, the stacking interactions are quite strong, with an ⁶⁷ interaction energy of -2.0 kcal/mol.²⁷

68 Our recent quantum chemical calculations²⁸ of pairs of 69 amyloid β -sheets indicate that the aromatic–aliphatic inter-70 actions contribute the most to amyloid stability, in the case of 71 amyloids with aromatic amino acids, while aliphatic–backbone 72 interactions contribute the most to amyloid stability in the case 73 of aliphatic amyloids (amyloids without aromatic residues). 74 These results also support previous findings that amyloid β -75 structures can be formed by nonaromatic peptides. Also, our 76 calculations of cyclohexane–benzene³¹ and benzene–ben-77 zene²⁷ interactions indicate stronger aromatic–aliphatic inter-78 actions than the aromatic–aromatic ones.

⁷⁹ In several studies, $\pi - \pi$ interactions in amyloids were ⁸⁰ detected through indirect experimental methods, like UV ⁸¹ fluorescence spectroscopy,²⁸ circular dichroism,^{11,30} and the ⁸² importance of these interactions in amyloid aggregations was ⁸³ assessed without any atomic level characterization.

In the present study, a systematic investigation of interactions of aromatic side chains at atomic resolution was performed by using X-ray and NMR structures of amyloids deposited in the PDB. Both natural and synthetic amyloids were included in the study, by taking only cross- β and parallel coil structures into account, in a thorough search of PDB for amyloids. Since fibril or growth can be conducted through β -sheet by increasing the hydrogen bonding between β -strands within the same β -sheet, or by side chain interactions between different β -sheets,¹⁰ these two types of Ar/Ar interactions were distinguished as intrasheet and intersheet (Figure 2). To the best of our knowledge, this is the first systematic study of interactions of aromatic side chains in all amyloid structures deposited in the PDB.



Figure 2. Example of intrasheet and intersheet contacts in an amyloid protein, PDB ID 2NNT.

97 MATERIALS AND METHODS

PDB Database. Amyloid protein three-dimensional (3D) 99 structures were searched in the PDB³¹ and in the CSD.³² The 100 searching criterion for the CSD was any at least four residue long 101 acyclic polypeptide with a nearly β-sheet structure. Eight structures 102 were found, but with no proof of self-assembly in the published papers. 103 An amyloid PDB subdatabase was made by searching the PDB for an 104 amyloid precursor name in the case of natural amyloids, and for the 105 terms *amyloidogenic, amyloid-related,* and *amyloid-like* for synthetic amyloids. Only the β secondary structures or coils were taken into 106 account. The details of the procedure for the database search have 107 been explained previously.³³ There were 109 structures found in the 108 PDB that fit these criteria, resolved by X-ray crystallography, solid state 109 or solution NMR. Some NMR structures are multiframe with up to 20 110 conformers, so total of 303 conformers were analyzed. There are 83 111 different peptide sequences in the constructed database. The X-ray 112 structures have been translated and rotated in order to obtain full 113 crystal lattice and biological assembly defined in PDB files; after, 114 duplicate interactions and amino acids have been excluded. In order to 115 determine the occurrence and impact of aromatic rings, all interactions 116 of aromatic rings, both aromatic–aromatic (Ar/Ar) and aromatic– 117 nonaromatic (Ar/nAr), were analyzed in every one of these sequences. 118

Aromatic–Aromatic Interactions. All the combinations of 119 interactions between the three aromatic amino acids, Phe, Tyr, and 120 Trp, were taken into account. Histidine was not taken into 121 consideration because it can be charged and thus screen more delicate 122 $\pi-\pi$ interaction. For Trp, we considered interactions of the six- 123 membered ring, while we did not consider interactions of the five- 124 membered ring. We determined the center–center distance between 125 the rings (*d*), the angle between ring planes (P_1/P_2), the normal 126 distance between ring planes (*R*), and the offset between ring centers 127 (*r*), shown in Figure 3. The distance (*R*) represents the normal 128 f3



Figure 3. Geometric parameters determined for each PDB amyloid structure: the center–center distance between rings (*d*), the angle between ring planes (P_1/P_2), the normal distance between planes (*R*), the offset between ring centers (*r*).

distance of the center of one ring (Ω) to its projection onto the plane 129 of the other ring (Ω_p) . The horizontal displacement (offset) *r* 130 represents the distance of the center of one ring (Ω') to the projection 131 of the center of the other ring onto the plane of the first ring (Ω_p) . For 132 (P_1/P_2) angles other than zero, there are two alternative and unequal 133 pairs of *R* and *r*; we have used the higher *R* and its corresponding *r* 134 value, as in ref 26. The distances between the $C\alpha$ atoms of two 135 interacting amino acids have been calculated as well. The scripts for 136 the search and for the PDB file parsing were written in Python 137 (http://www.python.org/) by using the MDAnalysis python library.³⁴ 138 Duplicate interactions have been recognized as having the same *d* 139 distance and excluded.

Aromatic molecules can form other types of interactions as well, 141 such as C–H/O and C–H/ π . A contact was considered C–H/O 142 interaction if the distance between a hydrogen atom from the C–H 143 group of an amino acid and an oxygen atom from another amino acid 144 was less than 2.9 Å and the C–H–O angle larger than 110°.^{35,36} The 145 geometrical criteria for the C–H/ π interaction were the distance 146 between the H atom and the center of phenyl ring is shorter than 3.5 147 Å, the angle between the C–H vector and the phenyl ring center is in 148 the range 110–180°, and the angle between the vector H atom— 149 center of the ring and the vector normal to the ring is smaller than 150 30°.³⁷

We also distinguished Ar/Ar interactions when the aromatic rings 152 pertain to parallel and antiparallel strand, by defining angle between 153 vectors $C-C\alpha$ for the two residues. When this vector was less than 154 90°, the strands were considered parallel.

Aromatic–Nonaromatic Interactions. To describe the Ar/nAr 156 interactions, the minimum distance between heavy atoms of two 157 interacting amino acids was calculated, taking into consideration side 158 chains only. The backbone interactions were not considered, as they 159

Table 1. Number and Percentages of Aromatic Amino Acids in Amyloid Sequences and Structures, and Their Involvement in Aromatic–Aromatic or Aromatic–Nonaromatic Interactions^a

		no. aromatics involved in Ar/Ar							Ar/Ar	no. aromatics involved in Ar/nAr						
	no. sequences with aromatics		no. aromatics in sequences		no. structures with aromatics		no. aromatics in structures		intersheet		intrasheet		intersheet		intrasheet	
Phe	38	45.8%	54	55.1%	48	44.0%	3505	57.1%	171	2.8%	2663	43.4%	2759	45.0%	387	6.3%
Tyr	32	38.6%	40	40.8%	49	45.0%	2351	38.3%	670	10.9%	1210	19.7%	723	11.8%	403	6.6%
Trp	3	3.6%	4	4.1%	5	4.6%	280	4.6%	61	1.0%	155	2.5%	164	2.7%	89	1.5%
total	56	67.5%		98	78	71.6%	6	136	902	14.7%	4028	65.6%	3646	59.4%	879	14.3%
^a Ar/Ar	= arot	natic_arc	matic i	interaction	s Ar/	nAr = aror	matic—no	onaromati	c intera	ctions Th	ere are 8	3 sequen	ces and 1	09 struct	ures in t	otal

Table 2. Number and Percentages of Structures and Interactions Involving Aromatic Amino Acids⁴

			no. str	uctures		no. interactions					
		int	ersheet	int	rasheet	inter	sheet	intrasheet			
Ar/Ar	PhePhe	7	6.4%	21	19.3%	10	2.4%	1492	64.6%		
	PheTyr	2	1.8%	0	0.0%	3	0.7%	0	0.0%		
	PheTrp	0	0.0%	1	0.9%	0	0.0%	4	0.2%		
	TyrTyr	14	12.8%	15	13.8%	397	96.8%	797	34.5%		
	TrpTrp	0	0.0%	1	0.9%	0	0.0%	16	0.7%		
	total	21	19.3%	33	30.3%	410		2309			
Ar/nAr	Phe	48	44.0%	33	30.3%	6902	72.4%	877	43.6%		
	Tyr	48	44.0%	31	28.4%	1827	19.2%	963	47.9%		
	Trp	4	3.7%	5	4.6%	810	8.5%	170	8.5%		
	total	78	71.6%	51	46.8%	95	539	20	010		

 a Ar/Ar = aromatic–aromatic interactions, Ar/nAr = aromatic–nonaromatic interactions. Total number of structures is 109, 78 of which contain aromatic amino acids.

160 are not specific to an amino acid. The minimum distance between 161 heavy atoms was limited to 5.0 Å in the search, as the sum of van der 162 Waals radii never exceeds this value, according to the CHARMM 163 parameters.³⁸ The interactions were discriminated as intersheet with 164 the $C\alpha$ – $C\alpha$ distance > 8 Å, and intrasheet with $C\alpha$ – $C\alpha$ distance < 6 165 Å, according to the results for the Ar/Ar. One interaction was counted 166 as one pair of residues.

167 **Number of Rings Involved in Interactions.** The number of 168 aromatic amino acids taking part in Ar/Ar or exclusively Ar/nAr 169 interactions was determined in every amyloid structure. An Ar/Ar 170 interaction was defined within the area that corresponds to the ellipse 171 (r = 7.0 Å and R = 6.0 Å) according to the results for the Ar/Ar search. 172 Ar/nAr interactions were defined as maximum heavy atom—heavy 173 atom distance up to 5.0 Å, according to the Ar/nAr search.

174 **RESULTS AND DISCUSSION**

175 We searched and analyzed interactions of aromatic side chains 176 in the subdatabase formed by amyloid structures deposited in 177 the PDB from June 2016. In the PDB, 83 sequences and 109



Figure 4. Normal distance (R) dependence on the offset values (r).



Figure 5. Example of the most frequent geometrical arrangement of the intrasheet interactions. PDBid: 4R0P, $P_1/P_2 = 0.0^\circ$, r = 3.57 Å.

amyloid structures were found, while 67.5% sequences and 178 71.6% structures contain aromatic amino acids (Table 1). 179 t1 These data show that amyloid structures can exist without 180 aromatic amino acids, as was observed previously.⁹ Moreover, 181 in a number of structures with aromatic amino acids, the Ar/Ar 182 interactions do not exist; among 109 structures, the Ar/Ar 183 interactions were observed only in 48 structures, and 184 specifically intesheet Ar/Ar interactions are present only in 185 21 structures (19.3%, Table 2). Hence, our data confirm that 186 t2 neither aromatic amino acids nor Ar/Ar interactions are crucial 187 for amyloid existence.

The analysis has been done separately for the amino acids 189 Phe, Tyr, and Trp, in order to detect different substituents 190 influences. The occurrence of aromatic amino acids among all 191 amino acids in amyloids is 3.92% for Phe, 2.90% for Tyr and 192 0.29% for Trp, which is very similar to the occurrences in 193 general protein sequences taken from Uniprot,³⁹ for Phe and 194 Tyr (3.93% and 2.94%, respectively), while occurrence for Trp 195 is larger in general protein sequence, 1.29%. 196

As was mentioned above, the intersheet and intrasheet 197 interactions were analyzed separately. The intrasheet Ar/Ar 198 interactions are far more frequent than the intersheet ones 199 (2309 over 410, Table 2), and also a higher number of rings is 200



Figure 6. Geometrical parameters for the intrasheet aromatic–aromatic interactions: (A) Distribution of the offset values (*r*), (B) distribution of the normal distances (*R*), (C) center–center distance distribution, (D) distribution of the angle between aromatic rings, and (E) distribution of the $C\alpha$ – $C\alpha$ distances.



Figure 7. Torsion angle *T* of an aromatic amino acid between the atoms C, $C\alpha$, $C\beta$, and $C\gamma$.

201 involved in the intrasheet interactions (4028 over 902, Table 202 1).

203 Considering intersheet interactions, the data in Table 1 show 204 that the number of aromatic amino acids involved in Ar/nAr interactions (3646) is larger than the number of aromatic $_{205}$ amino acids involved in Ar/Ar interactions (902). Also, the $_{206}$ number of Ar/nAr interactions (9539) is larger than the $_{207}$ number of Ar/Ar interactions (410), as data in Table 2 indicate. $_{208}$ On the other hand, for the intrasheet interactions, the Ar/Ar $_{209}$ (2309) are more preferred than the Ar/nAr ones (2010, Table $_{210}$ 2), and also a higher number of rings is involved in Ar/Ar $_{211}$ (4028) than in Ar/nAr (879, Table 1).

Aromatic—**Aromatic Interactions.** The common charac- $_{213}$ teristic of all the Ar/Ar interactions found in amyloid PDB $_{214}$ structures is that the normal distance between rings (*R*) $_{215}$ decreases as the offset value increases (*r*), as seen in Figure 4. $_{216 \text{ f4}}$ The database search yielded 3573 contacts found to be within $_{217}$

Article



Figure 8. P_1/P_2 angle-offset dependence for intrasheet aromaticaromatic interactions. Different amino acid pairs represented in various colors. (A) P_1/P_2 (offset) function for all intrasheet interactions. (B) P_1/P_2 (offset) function for intrasheet interactions not exposed to the solvent.



Figure 9. Intrasheet aromatic–aromatic interactions. (A) Type Phe– Phe exhibits higher offset values (right), PDB ID 2LMQ, r = 4.95 Å. (B) Type Tyr–Tyr exhibits lower offset values (right), PDB ID 2M5K, r = 2.30 Å. Besides intraseet $\pi - \pi$ interactions between aromatic rings, Phe and Tyr residues have additional interactions with the opposite amyloid sheet. Phenylalanines interact through their π -cloud with hydrophobic residues in the surrounding (Leu, C–H/ π interaction), while tyrosines form hydrogen bonds with the opposite sheet backbone through their –OH group, O–H–O angle 155.24°, O–O distance 3.41 Å.

218 the area that corresponds to the ellipse (r = 7.0 Å and R = 6.0 219 Å).

Some longer amyloid peptides exhibit the structure of a β -221 turn- β and look like U-shaped β -sheets. The aromatic rings 222 contained in these unstructured turns, as well as in the unstructured β -strand extremities, were not accounted for in 223 the interaction analysis, as they do not give rise to the cross- β 224 amyloid structure (Figure S1). In other words, only the 225 interacting cross- β fragments were analyzed in this study, and 226 hence we analyzed 2719 interactions. The geometric 227 parameters were analyzed separately for the intersheet and 228 the intrasheet Ar/Ar interactions. 229

Intrasheet Aromatic—**Aromatic Interactions.** When it 230 comes to the intrasheet arrangements, more than two aromatic 231 rings are stacked, and the rings are arranged in a nearly parallel 232 orientation, with varying offset values. The structure in Figure 5 233 f5 represents one typical example of these structural motifs. The 234 data on geometries of these interactions are given in Figure 6, 235 f6 where the characteristic angles between ring planes, P_1/P_2 , are 236 $0-5^{\circ}$ and the offset values (r) are in the range 2.5–5.0 Å. The 237 $C\alpha-C\alpha$ distance is in a small range, since it corresponds to the 238 intrasheet distances and is a general property of proteins, about 239 \sim 4.7 Å.¹⁰ As the distance between two amino acids is constant, 240 the variation in the center–center distance between rings (d) 241 and in the offset value (r) (Figure 6) is due to the change in the 242 torsion angle C–C α –C β –C γ , presented in Figure 7. 243 f7

In order to probe the influence of the ring type, the P_1/P_2 244 angle-offset dependence was separately shown for the four 245 systems, Phe–Phe, Phe–Trp, Tyr–Tyr, and Trp–Trp, which 246 were found in the interactions involving combinations of three 247 aromatic amino acids (Figure 8A). All systems, except Trp– 248 f8 Trp, show a tendency toward parallel interactions, and a large 249 range of offset values. The systems with tryptophan exhibit less 250 parallel geometries, which could be the consequence of a small 251 number of these interactions (Table 2). Namely, all the Trp– 252 Trp interactions were found in one 10-framed NMR structure, 253 and all tryptophans were positioned toward the water 254 environment with higher conformational freedom (Figure S2). 255

In the polar solvent environment, the dielectric constant is 256 higher than in the hydrophobic core of proteins, and the polar 257 interactions screen the delicate $\pi - \pi$ interactions. Hence, Figure 258 8B presents the P_1/P_2 angle-offset dependence when all 259 intrasheet interactions with aromatic rings that are close to 260 the water environment are excluded. The comparison of data in 261 Figure 8A,B shows that the interactions in the polar solvent 262 environment have high P_1/P_2 angles and can have high offset 263 values. The interactions in the hydrophobic core show 264 tendencies toward smaller P_1/P_2 angles (Figure 8B). One can 265 notice that only Phe–Phe and Tyr–Tyr interactions occur in 266 the hydrophobic core.

In comparison to tyrosine aromatic rings, the phenylalanine 268 rings demonstrate a higher tendency to form intrasheet Ar/Ar 269 interactions with a larger range of offset values and large range 270 of inter-ring angles (Figure 8B). Visual inspection of the 271 amyloid structures indicated that Phe rings are found nearly 272 parallel to the sheet plane, while the Tyr rings point toward the 273 opposite amyloid sheet, as presented in the examples in Figure 274 f9 9. Tyr possesses the -OH group and can form hydrogen bonds 275 f9 with the opposite sheet backbone (Figure 9B); this is the 276 reason why offset values for Tyr-Tyr interactions are in a 277 relatively small range, and they have a small angle P_1/P_1 (almost 278 parallel interactions, Figure 8). Hence, hydrogen bonds of 279 -OH group of tyrosine with the opposite backbone are 280 responsible for different geometries in Phe-Phe and Tyr-Tyr 281 contacts. 2.82

Phenylalanine residues that form interactions at large offsets $_{283}$ (3.5–5.0 Å) can form simultaneous interactions with ring faces. $_{284}$ It was previously demonstrated that high offsets in phenyl- $_{285}$

		inte	rsheet Ar/n	ıAr		intrasheet Ar/nAr								
	Phe		Tyr		Trp			Phe		Tyr		Trp		
Leu	2018	21.2%	154	1.6%	185	1.9%	Val	266	13.2%	314	15.6%	0	0.0%	
Ile	1556	16.3%	450	4.7%	0	0.0%	Ser	32	1.6%	338	16.8%	0	0.0%	
Val	759	8.0%	300	3.1%	33	0.3%	Glu	218	10.8%	128	6.4%	0	0.0%	
Ala	757	7.9%	77	0.8%	0	0.0%	Asp	48	2.4%	9	0.4%	116	5.8%	
Glu	585	6.1%	105	1.1%	126	1.3%	Leu	155	7.7%	17	0.8%	0	0.0%	
Asn	599	6.3%	173	1.8%	11	0.1%	Ala	83	4.1%	61	3.0%	1	0.0%	
Arg	174	1.8%	20	0.2%	294	3.1%	Thr	6	0.3%	39	1.9%	21	1.0%	
The most frequent interactions are represented. Total number of the interactions is 0520 for the intersheet and 2010 for the interactions														

Table 3. Number and Percentages of Intersheet and Intrasheet Aromatic-Nonaromatic Interactions between Different Aromatic and Nonaromatic Residues^a

The most frequent interactions are represented. Total number of the interactions is 9539 for the intersheet and 2010 for the intrasheet interactions.



Figure 10. (A) Antiparallel beta-sheet arrangement prevents the intrasheet aromatic-aromatic interactions, PDB ID 3MD4, and (B) parallel arrangement results in the intrasheet interactions, PDB ID 2BEG. Dashed lines represent the backbone hydrogen bonds, and red sticks represent the aromatic amino acids.

286 phenyl interactions are favorable in supramolecular structures, 287 since the π -cloud can simultaneously interact with other entities 288 in the vicinity.^{26,27} In amyloids, the simultaneous interactions of 289 the Phe ring are interactions with nonaromatic residues; the 290 most frequent are interactions with the leucine side chain (Table 3). 291

+3

In the example shown in Figure 9A, the Phe ring forms a 2.92 293 parallel interaction with another Phe ring at a large offset and simultaneously interacts with leucine (Figure 9A). In strands 294 with Tyr, the Tyr protrudes between the side chains of the 295 opposite sheet, due to its side chain forming hydrogen bond 296 with the opposite sheet (Figure 9B). This Tyr arrangement 297 contributes to the relatively small range of offset values for 298 Tyr–Tyr interactions (Figure 8). 299

In contrast to the intersheet interactions (see below) the 300 301 intrasheet aromatic contacts do not involve C-H/O and C-302 H/ π interactions, as there is no geometric condition for these interactions. Namely, $C-H/\pi$ are impossible with small inter-303 304 ring angles (Figure 8), while the C-H/O interactions of Tyr (only Tyr possesses oxygen) are not possible for the small 305 306 offsets observed in amyloid structures (Figure 8).

The interstrand hydrogen bonds of the backbone groups 307 308 stabilize individual beta-sheets, and they are stronger than $\pi - \pi$ 309 interactions.¹⁰ Thus, the intrasheet Ar/Ar interactions are 310 probably the consequence of the steric condition inside a

protein β -sheet, although interactions between aromatic rings 311 also contribute to the stabilization of a sheet. Examples in 312 Figure 10 show that the antiparallel beta-sheet arrangement 313 f10 prevents the intrasheet Ar/Ar interactions, while the parallel 314 arrangement results in the intrasheet interactions. In the 315 structures where intrasheet interactions are present, they are 316 always arranged as an array of rings. This arrangement also 317 maximizes the intrasheet backbone hydrogen bonds between 318 the parallel β -strands, because the strands are always aligned 319 along the entire length, Figure 10. Also, intrasheet Ar/Ar 320 interactions are not formed in every parallel structure (in 10 out 321 of 36 structures interactions not formed), even when rings are 322 aligned, which indicates poor importance of the intrasheet Ar/ 323 Ar interactions in amyloids. 324

Intersheet Aromatic-Aromatic Interactions. Histo- 325 grams with geometric data for intersheet Ar/Ar interactions 326 are given in Figure 11 and show significant differences between 327 fill the inter- and intrasheet interaction geometries (Figure 6). In 328 contrast to the case of intrasheet interactions, the intersheet 329 interactions have no parallel ring arrangements; the P_1/P_2 330 angles for most of the interactions have values in the range 331 $30-40^{\circ}$ (Figure 11). The histogram with offset values has two 332 sharp maxima, at 2.5-3.0 Å and 5.5-6.5 Å (Figure 11), while 333 the range of offset values for intrasheet interactions is 2.5-5.0 Å 334 (Figure 6). The intersheet interactions exhibit a somewhat 335 larger distance (d) than the intrasheet interactions, and the C α - 336 $C\alpha$ distance is much larger, since the rings from different sheets 337 are pointed toward each other. 338

The intersheet interactions are mostly pairwise, in contrast to 339 the intrasheet interactions, where there are several rings stacked 340 subsequently. In order to probe the influence of aromatic ring 341 type the angle P_1/P_2 —offset dependence was obtained 342 separately for Phe-Phe, Phe-Tyr, and Tyr-Tyr interactions, 343 the three types of the intersheet interactions that were found in 344 amyloid structures (Figure 12A). Like the intrasheet 345 f12 interactions, the mixed-type interactions are not common; 346 however, in the intersheet interactions the Tyr-Tyr 347 interactions prevail, while in the intrasheet the majority of 348 interactions are Phe-Phe (Table 2).

The intersheet Ar/Ar interactions have values of geometric 350 parameters over a large range, indicating a variety of interaction 351 geometries. This could be the consequence of a higher ring 352 steric freedom, as amyloid sheets are not as close as the strands 353 inside a sheet. The interactions of two aromatic rings with 354 inter-ring angles around zero and with offsets up to 2.0 Å had 355 been considered as stacking interactions. Recently, Zarić and 356 co-workers found significantly strong benzene-benzene 357 interactions also at larger offsets, up to 5.5 Å.^{26,27} Furthermore, 358 the interactions with P_1/P_2 angles up to 40° could be 359



Figure 11. Geometry parameters for the intersheet aromatic–aromatic interactions: (A) Distribution of the offset values (r), (B) distribution of the normal distances (R), (C) center–center distance distribution, (D) distribution of the angles between aromatic rings, and (E) distribution of the $C\alpha-C\alpha$ distances.

³⁶⁰ considered stacking, as they exhibit an energy-offset depend-³⁶¹ ence like that in the parallel interactions.²⁶

Most of the intermolecular Ar/Ar interactions are displaced 363 stackings (Figure 12B), although they can form other types of 364 interactions as well, like C–H/O^{36,37} and C–H/ π .³⁷ The 365 number of C–H/O interactions is 22, and the number of C– 366 H/ π interactions is 31 (Figure 12B). The structures in Figure 367 13 exemplify characteristic interactions between aromatic 368 moieties: stacking, displaced stacking, C–H/O and C–H/ π 369 interactions.

The rest, namely, the 176 "other" interactions, do not satisfy r1 criteria for any of the interactions. However, they are all are attractive. The potential surface for Ar/Ar interactions²⁰ shows that the interactions at offsets around 3.0 Å with P_1/P_2 angles 373 around 50° have interaction energies close to -2.0 kcal/mol, 374 while the interactions at offsets around 6.0 Å with P_1/P_2 angles 375 around 30° have interaction energies between -0.5 and -1.0 376 kcal/mol. The vast majority (145/176) of these "other" 377 interactions belong to a single 20-framed NMR structure, 378 PDB ID 2M5N, as shown in Figure S3. 379

The geometrical parameters for the interacting rings 380 belonging to the parallel and antiparallel strands are very 381 similar (Figure 12C), indicating that orientation of strands does 382 not have a significant influence on the intersheet interactions. 383 **Aromatic–Nonaromatic Interactions.** The intrasheet 384

Ar/nAr interactions are long with no peak at lower heavy 385



Figure 12. (A) P_1/P_2 angle-offset dependence for the intersheet aromatic–aromatic interactions. (B) Influence of the aromatic amino acid type. (C) Various types of interactions.



Figure 13. Representative structures of the intersheet interactions found in amyloids. (A) stacking: $P_1/P_2 = 10.90^\circ$, r = 1.94 Å, PDBid 4OLR, (B) displaced stacking: $P_1/P_2 = 33.78^\circ$, r = 3.02 Å, PDBid 2M5N, (C) C-H/ π interactions: $P_1/P_2 = 76.51^\circ$, r = 2.25 Å, PDBid 2NNT, (D) C-H/O interaction: $P_1/P_2 = 79.90^\circ$, r = 1.13 Å, PDBid 2NNT. The green dotted lines represent the putative interactions.





Article

Figure 14. Histogram of minimum distances between heavy atoms of two amino acids in aromatic–nonaromatic interactions; (A) intrasheet (B) intersheet.

These intrasheet interactions could also be the result of steric ³⁸⁹ conditions. The number of the interactions show that the ³⁹⁰ intrasheet Ar/nAr interactions are not particularly important ³⁹¹ (Table 1). ³⁹²

Differently than interacting distances of Ar/nAr intrasheet 393 interaction, the distances of intersheet interactions exhibit a 394 peak at 3.5-4.0 Å (Figure 14B). As the nonaromatic residues 395 are much more numerous (in average, every aromatic ring 396 interacts with \sim 3 nonaromatic, and \sim 1.5 aromatic residues), we 397 performed an analysis of the number of rings taking part in 398 certain types of interactions: Ar/Ar or Ar/nAr. Four times 399 higher number of aromatic rings takes part in the intersheet Ar/ $_{400}$ nAr than in the Ar/Ar interactions (3646 over 902, Table 1). 401 The interaction energy of Ar/nAr interaction can be also 402 substantial, comparable or even stronger than the Ar/Ar ones, 403 as shown by the interactions energy calculations.²⁸ Considering 404 particular amino acids, Phe has the highest preference toward 405 Ar/nAr interactions, while Tyr does not have large preference 406 for Ar/nAr interactions (Table 1). 407

The intersheet Ar/nAr interactions were found to involve 408 mostly aliphatic amino acids, especially Leu and Ile (24.7% and 409 21.0%, Table 3). An example of the interaction between Phe 410 and Leu is shown in Figure 9A.

Among the aromatic amino acids, phenylalanine was found 412 to be the most frequent in these contacts. 413

The greater impact of the intersheet Ar/nAr over Ar/Ar $_{414}$ interactions confirms previous experimental findings that $_{415}$ aromatic amino acid properties other than aromaticity could $_{416}$ be more important for amyloids, such as hydrophobicity, low $_{417}$ chain flexibility, and β -sheet propensity.^{18,21,40} 418

f14

419 **CONCLUSIONS**

420 By analyzing the aromatic—aromatic interactions in amyloids in 421 the PDB, it was established that aromatic amino acids are not 422 present in every amyloid sequence, and thus they are not 423 essential for amyloid self-assembly. The aromatic—aromatic 424 interactions in amyloids are less frequent than aromatic— 425 aliphatic interactions. In addition, the aromatic—aliphatic 426 interactions are present in more structures than the 427 aromatic—aromatic ones. Aromatic rings in amyloids tend 428 much more to interact with nonaromatic residues, especially 429 aliphatic ones, which is partially caused by a small number of 430 aromatic and a high number of aliphatic amino acids in the 431 amyloid sequences.

432 The aromatic—aromatic interactions between adjacent β -433 strands within the same β -sheet of an amyloid protein structure 434 are far more frequent than the intersheet interactions. Since the 435 aromatic—aromatic interactions are predominantly of the 436 intrasheet type, one can conclude that they play a less 437 dominant role for the association of amyloid sheets.

For the intrasheet aromatic–aromatic interactions, a parallel 439 displaced geometry is the most frequent, with the P_1/P_2 440 interplanar angles of 0–5° and varying offset values in the 441 range of 2.5–5.0 Å. In the case of the intersheet interactions, 442 there are no parallel ring arrangements. The most frequent are 443 displaced rings with P_1/P_2 interplanar angles between 30 and 444 40°.

445 **ASSOCIATED CONTENT**

446 Supporting Information

447 The Supporting Information is available free of charge on the 448 ACS Publications website at DOI: 10.1021/acs.cgd.7b01035.

- 449 Figure of an example of turn and coil aromatic amino
- 450 acids in amyloids (Figure S1), figure of the structure with
- 451 Trp-Trp intrasheet interactions (Figure S2), and figure
- 452 of the 20-framed NMR structure, PDBid 2M5N (Figure
- 453 S3) (PDF)

454 **AUTHOR INFORMATION**

455 Corresponding Author

- 456 *Phone: 011-3336605. E-mail: szaric@chem.bg.ac.rs.
- 457 ORCID (0)
- 458 Michael B. Hall: 0000-0003-3263-3219
- 459 Snežana D. Zarić: 0000-0002-6067-2349
- 460 Notes
- 461 The authors declare no competing financial interest.

462 **ACKNOWLEDGMENTS**

463 This work was supported by an NPRP grant from the Qatar
464 National Research Fund (a member of the Qatar Foundation)
465 [Grant Number NPRP8-425-1-087]. I.M.S. is grateful to the
466 Serbian Ministry of Education, Science and Technological
467 Development [Grant Number 172065] for supporting this
468 work.

469 **REFERENCES**

- 470 (1) Laurén, J.; Gimbel, D. A.; Nygaard, H. B.; Gilbert, J. W.; 471 Strittmatter, S. M. *Nature* **2009**, 457, 1128–1132.
- 472 (2) Haataja, L.; Gurlo, T.; Huang, C. J.; Butler, P. C. Endocr. Rev.
 473 2008, 29, 303–316.
- 474 (3) Ferreira, S. T.; Vieira, M. N. N.; De Felice, F. G. *IUBMB Life* 475 **2007**, *59*, 332–345.

- (4) Irvine, G. B.; El-Agnaf, O. M.; Shankar, G. M.; Walsh, D. M. *Mol.* 476 *Med.* **2008**, *14*, 451–464. 477
- (5) Murphy, M. P.; LeVine, H. J. Alzheimer's Dis. 2010, 19, 311–323. 478
 (6) Pulawski, W.; Ghoshdastider, U.; Andrisano, V.; Filipek, S. Appl. 479
 Biochem. Biotechnol. 2012, 166, 1626–1643. 480
- (7) Slotta, U.; Hess, S.; Spiess, K.; Stromer, T.; Serpell, L.; Scheibel, 481 T. *Macromol. Biosci.* **200**7, *7*, 183–188. 482
- (8) Li, C.; Mezzenga, R. Nanoscale 2013, 5, 6207–6218. 483
- (9) Lakshmanan, A.; Cheong, D. W.; Accardo, A.; Di Fabrizio, E.; 484 Riekel, C.; Hauser, C. A. E. *Proc. Natl. Acad. Sci. U. S. A.* **2013**, *110*, 485 519–524. 486
- (10) Harrison, R. S.; Sharpe, P. C.; Singh, Y.; Fairlie, D. P. Rev. 487 Physiol. Biochem. Pharmacol. **2007**, 159, 1–77. 488
- (11) Bowerman, C. J.; Liyanage, W.; Federation, A. J.; Nilsson, B. L. 489 Biomacromolecules **2011**, *12*, 2735–2745. 490
- (12) Cukalevski, R.; Boland, B.; Frohm, B.; Thulin, E.; Walsh, D.; 491 Linse, S. ACS Chem. Neurosci. **2012**, *3*, 1008–1016. 492
- (13) Doran, T. M.; Kamens, A. J.; Byrnes, N. K.; Nilsson, B. L. 493 Proteins: Struct., Funct., Genet. 2012, 80, 1053–1065. 494
- (14) Milardi, D.; Sciacca, M. F. M.; Pappalardo, M.; Grasso, D. M.; 495 La Rosa, C. *Eur. Biophys. J.* **2011**, 40, 1–12. 496
- (15) Tu, L.-H.; Raleigh, D. P. Biochemistry 2013, 52, 333–342. 497
- (16) Azriel, R.; Gazit, E. J. Biol. Chem. 2001, 276, 34156-34161. 498
- (17) Görbitz, C. H. Chem. Commun. 2006, No. No. 22, 2332-2334. 499
- (18) Lee, N. R.; Bowerman, C. J.; Nilsson, B. L. *Biomacromolecules* 500 2013, 14, 3267–3277. 501
- (19) Qi, R.; Luo, Y.; Ma, B.; Nussinov, R.; Wei, G. *Biomacromolecules* 502 2014, 15, 122–131. 503
- (20) Profit, A. A.; Vedad, J.; Saleh, M.; Desamero, R. Z. B. Arch. 504 Biochem. Biophys. 2015, 567, 46–58. 505
- (21) Bemporad, F.; Taddei, N.; Stefani, M.; Chiti, F. Protein Sci. 506 2006, 15, 862–870. 507
- (22) Profit, A. A.; Felsen, V.; Chinwong, J.; Mojica, E.-R. E.; 508
 Desamero, R. Z. B. Proteins: Struct., Funct., Genet. 2013, 81, 690-703. 509
 (23) Gazit, E. FASEB J. 2002, 16, 77-83. 510
- (24) Anjana, R.; Vaishnavi, M. K.; Sherlin, D.; Kumar, S. P.; Naveen, 511 K.; Kanth, P. S.; Sekar, K. *Bioinformation* **2012**, *8*, 1220–1224. 512
- (25) Chourasia, M.; Sastry, G. M.; Sastry, G. N. Int. J. Biol. Macromol. 513 2011, 48 (4), 540–552. 514
- (26) Ninković, D. B.; Andrić, J. M.; Malkov, S. N.; Zarić, S. D. Phys. 515 Chem. Chem. Phys. **2014**, *16*, 11173–11177. 516
- (27) Ninković, D. B.; Janjić, G. V.; Veljković, D. Ž.; Sredojević, D. N.; 517 Zarić, S. D. ChemPhysChem **2011**, *12*, 3511–3514. 518
- (28) Ninković, D. B.; Malenov, D. P.; Petrović, P. V.; Brothers, E. N.; 519 Niu, S.; Hall, M. B.; Belić, M. R.; Zarić, S. D. *Chem. - Eur. J.* **2017**, 23, 520 11046.
- (29) Ninković, D. B.; Vojislavljević-Vasilev, D. Z.; Medaković, V. B.; 522 Hall, M. B.; Brothers, E. N.; Zarić, S. D. Phys. Chem. Chem. Phys. **2016**, 523 18, 25791–25795. 524
- (30) Bowerman, C. J.; Ryan, D. M.; Nissan, D. A.; Nilsson, B. L. *Mol.* 525 *BioSyst.* **2009**, *5*, 1058–1069. 526
- (31) Berman, H. M.; Westbrook, J.; Feng, Z.; Gilliland, G.; Bhat, T. 527 N.; Weissig, H.; Shindyalov, I. N.; Bourne, P. E. *Nucleic Acids Res.* 528 **2000**, *28*, 235–242. 529
- (32) Allen, F. H. Acta Crystallogr., Sect. B: Struct. Sci. 2002, 58, 380- 530 388. 531
- (33) Stanković, I.; Hall, M. B.; Zarić, S. D. *Trans. Internet Res.* **2017**, 532 13. 533
- (34) Michaud-Agrawal, N.; Denning, E. J.; Woolf, T. B.; Beckstein, 534 O. J. Comput. Chem. 2011, 32, 2319–2327. 535
- (35) Dragelj, J. L.; Stanković, I. M.; Božinovski, D. M.; Meyer, T.; 536 Veljković, D. Z.; Medaković, V. B.; Knapp, E. W.; Zarić, S. D. *Cryst.* 537 *Growth Des.* **2016**, *16*, 1948–1957. 538
- (36) Veljković, D. Ž.; Janjić, G. V.; Zarić, S. D. *CrystEngComm* **2011**, 539 13, 5005. 540
- (37) Milčić, M. K.; Medaković, V. B.; Sredojević, D. N.; Juranić, N. 541 O.; Zarić, S. D. Inorg. Chem. **2006**, 45, 4755–4763. 542
- (38) MacKerell, A. D. J.; Bashford, D.; Bellott, M.; Dunbrack, R. L.; 543 Evanseck, J. D.; Field, M. J.; Fischer, S.; Gao, J.; Guo, H.; Ha, S.; 544

545 Joseph-McCarthy, D.; Kuchnir, L.; Kuczera, K.; Lau, F. T. K.; Mattos, 546 C.; Michnick, S.; Ngo, T.; Nguyen, D. T.; et al. *J. Phys. Chem. B* **1998**, 547 *102*, 3586–3616.

- 548 (39) Apweiler, R.; Bairoch, A.; Wu, C. H.; Barker, W. C.; 549 Boeckmann, B.; Ferro, S.; Gasteiger, E.; Huang, H.; Lopez, R.;
- 550 Magrane, M.; Martin, M. J.; Natale, D. A.; O'Donovan, C.; Redaschi, 551 N.; Yeh, L.-S. L. Nucleic Acids Res. **2004**, 32, 115D–119.
- 552 (40) Senguen, F. T.; Lee, N. R.; Gu, X.; Ryan, D. M.; Doran, T. M.;
- 553 Anderson, E. A.; Nilsson, B. L. Mol. BioSyst. 2011, 7, 486–496.